

# The complete genome of *Mycoplasmaoides cavipharyngis* strain 117C, a close relative of hemotrophic mycoplasma

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**ABSTRACT** *Mycoplasmaoides cavipharyngis* is one of the closest relatives of the highly host-adapted and uncultivable hemotrophic mycoplasma. The complete genome of strain 117C was constructed from long-reads derived from Pacific Biosciences single-molecule, real-time sequencing technology. The genome is organized into one circular, gapless chromosome with a length of 1,034 kb.

**KEYWORDS** complete genome, de novo, *Mycoplasmaoides cavipharyngis*

*Mycoplasmaoides* (synonym: [*Mycoplasma*]) *cavipharyngis* (1), initially isolated from the nasopharynx of guinea pigs, is characterized as a glucose-fermenting mycoplasma with a 96.5% identity to the 16S rRNA gene of *Mycoplasmaoides fastidiosum* (2, 3). Phylogenetic analyses based on 16S rRNA sequences revealed the formation of a distinct cluster for *M. cavipharyngis* and *M. fastidiosum* within the *Mycoplasmaoides pneumoniae* group, designated as the *M. fastidiosum* cluster (3). This cluster is described as being most closely related to the hemotrophic mycoplasma (HM) (3–5). HMs are highly specialized, hitherto uncultivable bacteria that cause infectious hemolytic anemia in various mammals worldwide (6). In contrast, *M. fastidiosum* and *M. cavipharyngis* are considered apathogenic species with established *in-vitro* cultivation systems (2, 7). Therefore, genome information on *M. cavipharyngis* and *M. fastidiosum* is crucial for understanding the evolutionary separation of HM from other mycoplasma species and for deciphering new approaches for an *in-vitro* cultivation system.

*M. cavipharyngis* strain 117C [ATCC 43016] was cultivated in SP-4 medium (ATCC Medium 988, [8]) at 37°C under 5% CO<sub>2</sub>. DNA extraction was performed using the Monarch HMW DNA Extraction Kit (NEB, Frankfurt, Germany), followed by DNA quantification with a Qubit Fluorometer (Thermo Fisher Scientific, Darmstadt, Germany). NEB ultra-long reads were obtained through PacBio single-molecule, real-time (SMRT)-sequencing performed at the Max-Planck-Genome-Centre (Cologne, Germany) on a PacBio Sequel II device using SMRTbell Express Template Preparation Kit 2.0, along with Binding Kit 2.0, and Sequel II Sequencing Kit 2.0 (PacBio, CA, USA). The Sequencing resulted in 91,189 reads with an average length of 10,950 nt. Correction, trimming, and assembly of the achieved reads were performed using Canu v2.2 (9) with the pacbio-hifi option and an expected genome size of 1.0 Mb. Of the 91,189 reads, 18,229 corrected-trimmed reads ( $N_{50}$  of 12,906) were used for assembly, resulting in a single, circular, gapless contig of 1,065,917 bp in length and 186.63-fold coverage.

Sequence overlap was confirmed by BLAST+v.2.12.0 analysis (10) and removed using Artemis v18.2.0 (11). The final assembled genome was automatically annotated using RAST v2.0 (12) with the chromosome start set 21 bp prior to the *dnaA*. Annotation validation and curation were conducted manually in Artemis v18.2.0 (11) using BlastKOALA v3.0 (13), InterProScan v99.0 (14), and BLAST analysis against the non-redundant protein database (<https://www.ncbi.nlm.nih.gov/refseq/>). Missing non-coding RNA and ribosomal proteins were added using CMsearch v1.1.4 (15) and BLAST analysis.

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Quality control and completeness of the chromosome were assessed based on BUSCO v5.5.0 analysis (16) with 171 single-copy orthologs of *Mycoplasmatales* spp. (94.8%). All programs and tools used were run with default settings unless otherwise specified.

The circular genome of *M. cavipharyngis* consisted of 1,034,253 bp with a GC content of 25.28%. The rRNA genes were organized in one contiguous operon and a complete set of tRNAs was present. The coding sequences comprised 768 protein-coding genes and 18 pseudogenes.

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Janina Kramer, Data curation, Formal analysis, Investigation, Methodology, Project administration, Software, Validation, Writing – original draft, Writing – review and editing | Christina Zübert, Formal analysis, Methodology, Resources, Writing – review and editing | Bruno Huettel, Data curation, Formal analysis, Methodology, Resources, Software, Writing – review and editing | Michael Kube, Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Resources, Software, Supervision, Validation, Writing – review and editing | Ludwig E. Hoelzle, Conceptualization, Investigation, Resources, Supervision, Writing – review and editing

## DATA AVAILABILITY

Raw reads were allocated to Sequence Read Archive under accession number [SRR28961922](https://www.ncbi.nlm.nih.gov/sra/SRR28961922). A whole-genome project has been submitted to GenBank under BioProject accession number [PRJNA1107253](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1107253). The annotation has been deposited under accession number [CP155569.1](https://www.ncbi.nlm.nih.gov/CP155569.1).

## REFERENCES

- Gupta RS, Sawhani S, Adeolu M, Alnajjar S, Oren A. 2018. Phylogenetic framework for the phylum Tenericutes based on genome sequence data: proposal for the creation of a new order *Mycoplasmoidales* ord. nov., containing two new families *Mycoplasmoidaceae* fam. nov. and *Metamycoplasmataceae* fam. nov. harbouring *Eperythrozoon*, *Ureaplasma* and five novel genera. *Antonie Van Leeuwenhoek* 111:1583–1630. <https://doi.org/10.1007/s10482-018-1047-3>
- Hill AC. 1984. *Mycoplasma cavipharyngis*, a new species isolated from the nasopharynx of guinea-pigs. *J Gen Microbiol* 130:3183–3188. <https://doi.org/10.1099/00221287-130-12-3183>
- Johansson KE, Tully JG, Bölske G, Pettersson B. 1999. *Mycoplasma cavipharyngis* and *Mycoplasma fastidiosum*, the closest relatives to *Eperythrozoon* spp. and *Haemobartonella* spp. *FEMS Microbiol Lett* 174:321–326. <https://doi.org/10.1111/j.1574-6968.1999.tb13585.x>
- Tasker S, Helps CR, Day MJ, Harbour DA, Shaw SE, Harrus S, Baneth G, Lobetti RG, Malik R, Beauflis JP, Belford CR, Gruffydd-Jones TJ. 2003. Phylogenetic analysis of hemoplasma species: an International Study. *J Clin Microbiol* 41:3877–3880. <https://doi.org/10.1128/JCM.41.8.3877-3880.2003>
- Peters IR, Helps CR, McAuliffe L, Neimark H, Lappin MR, Gruffydd-Jones TJ, Day MJ, Hoelzle LE, Willi B, Meli M, Hofmann-Lehmann R, Tasker S. 2008. RNase P RNA gene (*rnpB*) phylogeny of hemoplasmas and other *Mycoplasma* species. *J Clin Microbiol* 46:1873–1877. <https://doi.org/10.1128/JCM.01859-07>
- Messick JB. 2004. Hemotrophic mycoplasmas (hemoplasmas): a review and new insights into pathogenic potential. *Vet Clin Pathol* 33:2–13. <https://doi.org/10.1111/j.1939-165x.2004.tb00342.x>
- Lemcke RM, Poland J. 1980. *Mycoplasma fastidiosum*: a new species from horses. *Int J Syst Bacteriol* 30:151–162. <https://doi.org/10.1099/00207713-30-1-151>
- Tully JG. 1995. Culture medium formulation for primary isolation and maintenance of mollicutes, p 33–40. In Razin S (ed), *In Molecular and Diagnostic Procedures in Mycoplasma*. Elsevier professional.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive *k*-mer weighting and repeat separation. *Genome Res* 27:722–736. <https://doi.org/10.1101/gr.215087.116>

10. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
11. Rutherford K, Parkhill J, Crook J, Horsnell T, Rice P, Rajandream M-A, Barrell B. 2000. Artemis: sequence visualization and annotation. *Bioinformatics* 16:944–945. <https://doi.org/10.1093/bioinformatics/16.10.944>
12. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, et al. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>
13. Kanehisa M, Sato Y, Morishima K. 2016. BlastKOALA and GhostKOALA: KEGG tools for functional characterization of genome and metagenome sequences. *J Mol Biol* 428:726–731. <https://doi.org/10.1016/j.jmb.2015.11.006>
14. Jones P, Binns D, Chang H-Y, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong S-Y, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>
15. Nawrocki EP, Eddy SR. 2013. Infernal 1.1: 100-fold faster RNA homology searches. *Bioinformatics* 29:2933–2935. <https://doi.org/10.1093/bioinformatics/btt509>
16. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>